



REVIEW ARTICLE

Hairy cell leukemia, an uncommon B-cell lymphoid neoplasia



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Abstract Hairy cell leukemia (HCL) is an uncommon B-cell lymphoid neoplasia, representing 2–3% of all leukemias. It is more common in men, with a median age at diagnosis of 52 years. The hair-like projections on its surface are its principal characteristic. Although its etiology has not been established, the recent gene sequencing of HCL identified the presence of the BRAF V600E mutation, absent in other malignant neoplasias of the lymph cells. The clinical course of the disease is usually indolent. The majority of patients initially present weakness and fatigue, pancytopenia and splenomegaly.

HCL must be distinguished from other indolent lymphoid neoplasias such as prolymphocytic leukemia, splenic marginal zone lymphoma, the variant of HCL (HCLv), and mantle cell lymphoma. Peripheral blood flow cytometer is essential for the detection of CD11c, CD19, CD20, CD22, CD25, and CD10, as well as a bone marrow aspirate to detect immunophenotypes. In addition, a bone biopsy is useful to perform an immunohistochemistry analysis for TRAP, DBA-44 and A1 annexin.

Purine analogues remain the first line of treatment. However, interferon and rituximab are a valid option, if the ideal treatment is unavailable. New discoveries in the pathophysiology of HCL have brought the creation of pharmaceuticals with distinct therapeutic targets. These pharmaceuticals are currently undergoing testing.

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Introduction

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Hairy cell leukemia (HCL) is a B-cell lymphoid neoplasia. HCL differentiates from other B-cell neoplasms because of its morphological, immunophenotypic and molecular characteristics. Its main characteristic is the accumulation of monoclonal B cells, with hair-like projections on its

surface, mainly found in peripheral blood, bone marrow and the spleen. It was described in 1958 by Bournoncle et al. However, they called it leukemic reticuloendotheliosis.¹ The term "hairy cells" was coined in 1996 by Schrek et al.² Today it is classified by the World Health Organization as a B-cell non-Hodgkin lymphoma.³

Definition

HCL is a rare, chronic, lymphoproliferative B-cells disorder. Its main characteristics are the cytoplasmic prolongations which give the cells a hairy aspect.

Etiology

Although there are different studies showing a link between exposure to certain agents and HCL, its etiology has not been clearly established, which could be explained due to its low incidence. Amongst the agents which have proven a positive link are: exposition to pesticides,⁴ herbicides, mineral oils, working as a carpenter, or a farmer.⁴⁻⁷ Recently, a positive link with size has been described,⁵ whereas smoking has an inverse association, especially in men.⁸ The specific mechanism which confers the protection is unknown; nevertheless, smoking has been suggested to decrease the severity of inflammatory mechanisms⁹ and that nicotine induces apoptosis in lymphocytes.¹⁰

Epidemiology

HCL represents 2–3% of all leukemias. There are 600 new cases reported per year in the U.S. (3.2 cases per million inhabitants). Median age at diagnosis is 52 years old and it is more common in men than in women, with a 4:1 ratio; with a higher incidence in the white population, especially among Ashkenazi Jews.³ In Mexico, HCL represents 1.12% of all leukemias. However, in the north of the country it represents up to 1.83%, similar to the information from the U.S.^{11,12}

Physiopathology

HCL is a chronic, lymphoproliferative B-cell disorder. However, its cells do not have the appearance of any B-cell sub-population, and its origin has been a matter of debate. The analysis of immunoglobulins (Ig) variable regions genes is a tool used to discover the lymphoid cells' clonal origin.¹³ In over 85% of cases, we are able to find somatic mutations in Ig variable regions genes of HCL cells,^{14,15} which is an indication that the cells involved have passed through the germinal center or lymphoid peripheral organs.¹⁶ Approximately 40% of HCL cells co-express multiple clonally-related Ig isotypes.¹⁷

Evidence suggests an origin post germinal center in memory B-cells, due to their genomic expression profile.^{18,19} The origin of memory B-cells is compatible with HCL's lack of chromosomal reciprocal translocation.²⁰ Absence of CD27 is typical in HCL. This represents a point against the hypothesis of its origin in memory B-cells.²¹ However, negative CD27 memory B-cells have been observed.²² Since lymph node

affection is infrequent, it has been suggested that the HCL origin cell is probably located in the bone marrow or spleen, since those are usually the more affected places. HCL cells show an expression profile similar to the spleen's marginal zone.²³

HCL's characteristic look is due to the beta-actin expression, which is polymerized to F-actin, located in the cortical cytoskeleton.²⁴ PP52 phosphoprotein, which is specific for leukocytes, is linked to F-actin and is responsible for bringing support to the hair-like projections.²⁵ On the other hand, HCL gene sequencing recently identified the presence of a BRAF V600E mutation in almost every patient with the disease, absent in other B-cell lymphoid malignancies.^{26,27} BRAF mutations activate the MAPK pathway, promoting growth, survival and HCL cell differentiation.²⁸

Clinical presentation and lab findings

The disease's clinical course is indolent. Most patients usually present weakness and fatigue as the predominant symptoms during the onset of the disease.²⁹ Sometimes, there is a history of repeated infections. The fysical examination findings are: splenomegaly in 96%, hepatomegaly in 58% and lymphadenopathy in only 35%. These swollen lymph nodes are rarely observed in the periphery; nonetheless, they are generally present in the abdomen and detected by imaging studies.³⁰

In an advanced stage of the disease we are able to find pain in the superior left quadrant, infections, fever, and hemorrhages and/or weight loss. However, this is uncommon because of the availability and effectiveness of treatment.^{30,31} Clinical manifestations are product of hairy cell accumulation in the spleen, liver and bone marrow (Table 1).³²

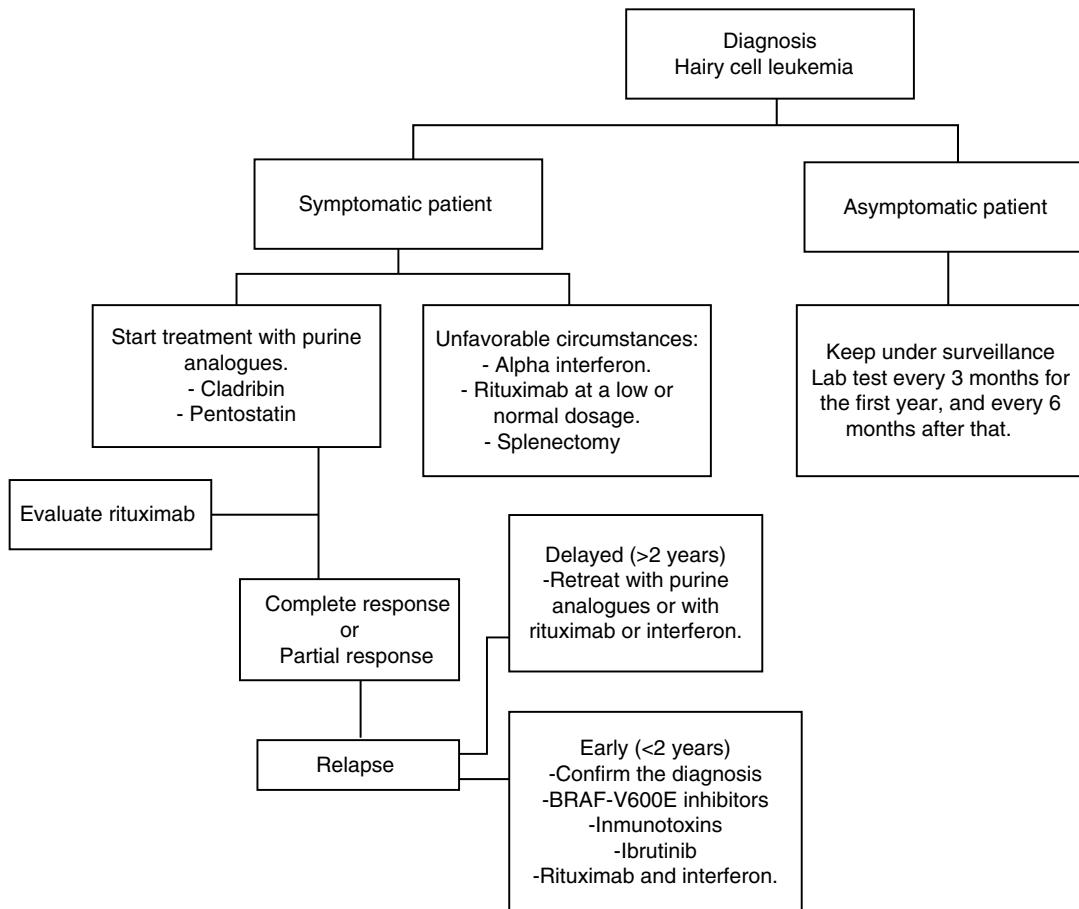
Respect to lab studies, it is frequent to observe anemia in 85%, thrombocytopenia in 60.80% and leukopenia in 60% due to hypersplenism and bone marrow infiltration.³⁰

Differential diagnosis

HCL must be differentiated from other indolent lymphoid malignancies such as prolymphocytic leukemia, splenic marginal zone lymphoma, mantle cell lymphoma and HCL variant (HCL-v). The last one occurs in 10% of cases with a median age of 70 years, and despite the similarities with classic hairy cell leukemia, they diverge in the absence of the CD25 and CD123 immunophenotypic markers. Another way of making a differential diagnosis is the lack of response to standard HCL treatment and the nonexistence of mutations of the BRAF V600F gene.³⁰

Diagnostic methods

HCL diagnosis is commonly made with a biopsy and bone marrow aspiration combined with immunophenotypic characterization through flow cytometry.³³ It is important to point out that this pathology is usually sub-diagnosed and requires clinical suspicion and the use of the proper technology to solve this problem. As previously mentioned, most patients (70–90%) present pancytopenia, with leucopenia ($<5 \times 10^9/L$), anemia ($<10\text{ g/dL}$), neutropenia



Algorithm 1 Chart for the management of a patient with hairy cell leukemia.

Table 1 Clinical manifestations.

Spleen	Liver	Bone marrow	Lymph nodes
The cells accumulate in the red pulp and atrophy the white pulp. They later form the so-called "pseudo-sinusoids" through the replacement of the endothelial cells of the red pulp by enhanced vascular channels, contributing to anemia.	Here, they accumulate in the liver sinusoids as well as in the portal tract. There is fibrosis in the latter due to the abundant hyaluronic acid, which stimulates the hairy cells to produce fibronectin, with its corresponding fibrosis.	In this area, there is extensive production of fibrosis and suppression of hematopoiesis. The key factor is the interaction of the hairy cells with the hyaluronic acid of the extracellular matrix, generating fibroblastic growth factors (FGF) and stimulating the malign cells to produce and secrete fibronectin.	Generally, absent of the disease. Lack of receptors for entry of hairy cells.

($<1 \times 10^9/L$), moncytopenia ($<0.1 \times 10^9/L$) and thrombocytopenia ($<100 \times 10^9/L$). Only between 10% and 20% present moderate leukocytosis ($>10 \times 10^9/L$). HCL patients display elevated IL-2R (CD25) serum levels, which correlates with the disease's degree of activity.³⁴ Other tests which should be considered when making the diagnosis are serum immunoglobulin levels, as well as IgVH gene and BRAF

V600E somatic mutations.³² Some HCL histopathologic and immunophenotypic characteristics are the following:

- Lymphocyte flow cytometry in peripheral blood or bone marrow with CD19, CD20, FMC7, CD11c, CD103, CD25, HC2, CD22, sIg, CD79a and CD123 expressions, with four being the main and specific markers: CD11c, CD103, CD25

Table 2 Criteria to begin treatment.

1. *Symptomatic splenomegaly*
2. *Cytopenia involving at least one of the following:*
 - Hemoglobin <10 g/dL
 - Platelets <100 × 10⁹/L
 - Neutrophils <1 × 10⁹/L
3. *Severe infections*

and CD123.³⁴ Commonly negative markers are CD5, CD23, CD10, CD79b and CD27.³²

- A strong expression for CD200 is characteristic of HCL and may be useful in the diagnosis of difficult cases.³⁴
- Bone marrow aspiration with a needle may be difficult to obtain and is frequently unproductive or "dry". In a bone marrow biopsy, we can observe fibrosis, with a cellular "fried-egg" look caused by the wide spaces between nuclei and abundant cytoplasm. Immunohistochemistry analyses for CD20 and TRAP (tartrate resistant acid phosphatase), DBA-4 and annexin A1 are conducted, which are characteristically positive.³²

Andrulis et al. directed a study where the efficiency of the VE1 antibody for BRAF V600E detection was reported, along with HCL identification in other entities. Moreover, a study conducted by Uppal et al. found a sensitivity of 88% and a specificity of 97% to detect this mutation with the mentioned antibody.³⁴

Current treatment

HCL regularly has an indolent evolution. Waiting and observing is a good option for asymptomatic patients, since early treatment does not offer any sort of benefit in the survival rates of these cases. Anyway, the progression of the disease in most patients will lead to complications as a result of cytopenias and splenomegaly; i.e. anemia, hemorrhages, recurrent infections, etc. In general practice, treatment must be started if any of the criteria listed in **Table 2** occur.³⁵ When the decision to not start treatment

is taken, clinical and laboratory monitoring should be done for every three months during the first year and every six months thereafter.³⁵ HCL treatment is not considered healing; but current treatment strategies are capable of reaching prolonged remissions, thus increasing global survival rates (**Algorithm 1**).³⁶⁻³⁹

Purine analogues

In 1960, findings showed that 30% of children with severe combined immunodeficiency syndrome lacked deaminase adenosine enzyme (ADA).⁴⁰ Furthermore, they discovered that the accumulation of the deoxyadenosine triphosphate form was responsible for the decrease of lymphocytes.⁴¹ Keeping these observations in mind, medications capable of binding irreversibly to ADA or antagonizing its action were developed. After treatment with purine analogues, deoxyadenosine triphosphate accumulation results in rupture and inhibition of the DNA repair, which translates into cellular apoptosis.⁴²

Pentostatin

Also known as 2'-deoxycoformycin (dcF), a product of *Streptomyces antibioticus* and ADA inhibitors, first introduced in 1980 as the first purine analogues for HCL treatment. In **Table 3**,^{37,39,43,44} specific studies show response rates to pentostatin. Intravenous use of pentostatin at a 4 mg/m² ratio once every two weeks until reaching complete response was approved in the U.S. Pentostatin is safe in patients with creatinine lightening >60 mL/min. However, a dosage reduction is necessary if this depuration is between 40 and 60 mL/min.³² Patient hydration with 1.5 L of intravenous solution with every pentostatin cycle is recommended.³⁰

Global response varies between 88% and 96%, while complete response is between 44% and 81%. Flinn et al. estimated a survival rate of 5 and 10 years of 90% and 81%, respectively, with a mean follow-up duration of 9.3 years.⁴⁵ Pentostatin is generally well-tolerated and the most common adverse effects are anemia, thrombocytopenia and neutropenia.⁴⁶ Still, pentostatin has been reported to

Table 3 Studies on the effectiveness of pentostatin in HCL cases.

Study	Follow-up	Plan	% CR	% PR	% GR	Outcome
Rafel et al. (n = 78)	2.6 years	IV infusion 4 mg/m ² /d Every 2 weeks	72	16	88	Event-free survival: 46 months
Else et al. (n = 185)	10.8 years	IV infusion 4 mg/m ² /d Every 2 weeks	81	15	96	Global survival at 10 years: 98%
Ribeiro et al. (n = 49)	3.2 years	IV infusion 4 mg/m ² /d Every 2 weeks	44	52	96	Global survival: 86%
Maloisel et al. (n = 238)	5.3 years	IV infusion 4 mg/m ² /d Every 2 weeks	79	17	96	Global survival at 5 years: 88.1% Global survival at 10 years: 68.8%

RC: complete response, RP: partial response, RG: global response.

Table 4 Studies on the effectiveness of cladribine in HCL cases.

Study	Patients	Follow-up	Plan	% CR	% PR	% GR	Outcome
Robak (1999)	97	36 months	2 h infusion IV 0.12 mg/kg/d 5 days	77.3	18.6	95.9	Progression-free survival: 37.4 months
Rosenberg (2014)	83	NA	Continuous infusion IV 0.1 mg/kg/d 7 days	88	12	100	Average global survival: 231 months
Goodman (2003)	207	7 years	Continuous infusion IV 0.1 mg/kg/d 7 days	95	5	100	Global survival at 108 months: 97%
Chadha (2005)	86	9.7 years	Continuous infusion IV 0.1 mg/kg/d 7 days	79	21	100	Global survival at 12 years: 87%

RC: complete response, RP: partial response, RG: global response.

decline CD4+ and CD8+ lymphocyte counts significantly, which could increase secondary malignancies and incidence of infections.^{47,48}

Cladribine

It is known as 2-chlorodeoxyadenosine (CdA). In Table 4,^{49–52} studies indicate its effectiveness. The most used scheme consists of administering 0.1 mg/kg/day in a continuous infusion for 7 days. In a non-randomized study, there was evidence proving that there was no statistically significant difference in the response and toxicity ranges between infusions (24 h and 2 h).⁵³ Another randomized study compared daily versus weekly cladribine administration; there were no significant findings in response, survival, global and toxicity rates.⁵⁴ A different study showed that the weekly program reduced the risk of infections.⁵⁵ One of the advantages of subcutaneous administration is the fact that in most cases it does not require hospitalization. 0.14 mg/day for 5 days has a 95% response rate,⁵⁶ similar to the intravenous administration. Weekly subcutaneous programs have similar response and toxicity rates as daily ones.⁵⁷ With just one cladribine cycle, a global response up to 100% can be obtained, and total response rates differ from 77% to 95%.^{49–52} Juhn et al. reported a global survival at 12 years of 79%.³⁶ In general, cladribine is well-tolerated, with cytopenias and fever being the most common adverse effects.

Until now, there are no randomized prospective studies comparing pentostatin versus cladribine, in part because of the great efficiency of both drugs and due to low HCL incidence. Even so, there are retrospective studies proving the fact that both drugs have a similar efficacy, in terms of complete response and free-of-disease survival.

Other treatments

Purine analogues remain the first line of treatment but new discoveries regarding HCL pathophysiology have led to the creation of drugs with different therapeutic targets. These drugs are under research and some have shown promising results.

Rituximab

Since HCL is a B-cell malignancy, it is logical to use a monoclonal antibody against CD20, such as rituximab. Employed as a stand-alone drug, rituximab may reach total response rates of 10–54% in patients with an HCL relapse, at a 375 mg/m² dosage once a week for 4–8 weeks.^{58,59} Else et al. retrospectively reviewed 18 patients who were treated with purine analogues in combination with rituximab as a second line of treatment, after being treated with purine analogues as single agents. All patients responded, with a complete response rate of 89%.⁶⁰

Rituximab at 375 mg/m² per week for 8 weeks as initial therapy after administrating 5.6 mg/m² cladribine via 2-h IV infusion for 5 days generates a total response of 100%.⁶¹ In special or unfavorable situations, 100 mg of rituximab per week may be utilized for 4–6 weeks. This is less expensive and is usually effective, especially when is combined with interferon.

While purine analogues may not be capable of eliminating HCL, since minimal residual disease (MRD) detected after cladribine administration is always strongly CD20+, MRD eradication may be obtained using rituximab. Rivandi et al. proved in a preliminary study that rituximab at conventional doses for a period of 8 weeks performs with great activity, eliminating MRD in 13 patients, when it is used 4 weeks after the administration of cladribine.⁶²

Vemurafenib

As described earlier, BRAF V600E mutation is the genetic key in HCL. Therefore, it is a therapeutic target which has been studied in recent years. Vemurafenib is a BRAF V600E oral inhibitor. Tacci et al. conducted a study to measure Vemurafenib activity and safety in patients with HCL who relapsed after treatment with purine analogues or who were refractory to the administration of purine analogues. Global response rate was 96% and complete response rate was 35%, with a relapse-free survival mean of 19 months. The adverse effects were rash, arthralgia and arthritis.⁶³

Since a positive relation has been observed between the use of Vemurafenib and the onset of dermatological

malignancies, frequent explorations of the skin are recommended.

Ibrutinib

A selective and irreversible inhibitor of the Burton's tyrosine kinase intervenes in the B-cell signaling pathway.⁶⁴ An ibrutinib clinical assay in patients with HCL relapse has recently begun. Preliminary efficacy and safety data show adverse effects such as eruptions, diarrhea and arthralgia. This clinical assay is currently taking place in several centers in the U.S. (NCT01841723).

Immunotoxins

In order to increase monoclonal antibody cytotoxicity, techniques that facilitate the production of antibody-toxin or antibody-drug conjugates have been created. An immunotoxin is the fusion between a bacterial toxin (i.e. *Pseudomonas exotoxin* or diphtheria) and the variable fraction of a monoclonal antibody, whose specific target is found on the surface of neoplastic cells like CD25 or CD22. This toxin is released in the neoplastic cell's interior and interferes with protein synthesis.⁶⁵

BL22 is an immunotoxin against CD22 fused with a truncated shape of the *P. exotoxin* PE38. In a phase II clinical assay, BL22 was tested in 36 HCL relapse or refractory disease cases. After a cycle (40 mg/kg every two days, three doses), complete response rate was 25% and global response rate was 50%. These responses improved to a total response rate of 47% and a global response rate of 72% after retreatment (only in patients with cytopenias). Two patients developed uremic hemolytic syndrome without the need to recur to plasmapheresis.⁶⁶ Subsequently, moxetumomab pasudotox was developed as a modified version of BL22 with a higher affinity and cytotoxicity. In a phase I assay, which included 28 patients with HCL relapse and resistance, a global response rate of 86% was obtained, including a sustained complete response rate in 46% of patients.⁶⁷

Therapeutic options in unfavorable circumstances

Even though HCL is treated in most developed countries with cladribine and pentostatin, it is a fact that these drugs are not only expensive, they are not available in Mexico and in many countries with limited resources. So, in these types of circumstances, there are other affordable therapeutic options with favorable results.

Interferon alpha for HCL patient treatment was first introduced in 1984. Today, its use is limited, mainly due to purine analogues' great effectiveness. On the other hand, in countries with low economic resources, it is an inexpensive option and one which has proved similar results to those of cladribine in terms of global survival. Ruiz-Delgado et al. conducted a comparative study between interferon alpha ($n=18$) and cladribine ($n=11$), where the difference in global survival between both groups was not statistically significant; 94% at 217 months in the interferon group and 91% at 133 months in the cladribine group.⁶⁸ In a study

conducted in our center, nine HCL patients received three IFN mega-units three times a week for 12 weeks, subsequently they received treatment once again for 8 weeks when there was a leukemia reactivation or after 10 months of observation each year. All patients had a hematological remission before 12 weeks of treatment. This therapeutic option is cheaper, effective and comparable to other forms of therapy with IFN in the treatment and maintenance of patients with this type of leukemia.⁶⁹ It is possible to combine interferon with rituximab without increasing toxic effects and improving effectiveness.

Splenectomy was the first intervention that significantly changed the survival in patients. Today, it is rarely used. It may be recommended in patients with painful massive splenomegaly (>10 cm under the costal edge) and with minimal infiltration of the bone marrow, or in patients refractory to treatment with interferon and purine analogues.³³ Retrospective studies show a complete response rate of 40–62%, and a mean survival rate at 5 years up to 68%.^{70,71} Lad et al. published a retrospective study, including 24 patients with an HCL diagnosis, they were divided into two groups: 17 patients received cladribine and 7 were splenectomized. 75% of patients in the splenectomy group showed total remission, 94% did so in the cladribine group. An interesting finding when comparing both groups was that no statistically significant differences were observed regarding leukemia-free survival and global survival.⁷²

Prognosis

Survival time in patients after diagnosis was 4 years before a treatment was known, due to complications derived from cytopenias, including hemorrhages and infections. Thereafter, with splenectomy as a first line treatment, there was a complete response of 40–62% and a survival rate of 5 years at 61–68%. Then, alfa interferon was utilized as the first drug with benefits in the treatment of HCL. Still its full response rate was low, at 10%.⁷³

Today, with purine analogues (pentostatin and cladribine), complete response is induced up to 80% of patients with a median survival of 10 years. Global response rates are 96–100% with a complete response rate of 80% and a survival rate of 10 years ranging between 85% and 100%.⁷⁴ Despite that, a significant proportion of patients with HCL fail in their response to treatment or become resistant. Up to 48% of patients relapse in the following 15 years.⁷⁵ The future of HCL patients is very favorable. The challenge is to identify this malignancy as early as possible and treat it properly using available resources.

Conflict of interest

The authors have no conflicts of interest to declare.

References

- Bouroncle BA. Leukemic reticuloendotheliosis (hairy cell leukemia). *Blood*. 1979;53:412–36.
- Schrek R, Donnelly W. "Hairy" cells in blood in lymphoreticular disease and "flagellated" cells of normal lymph nodes. *Blood*. 1966;27:199–211.

3. Swerdlow SH, Campo E, Harris NL, et al. WHO classification of tumours of haematopoietic and lymphoid tissues. Lyon, France: World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissue; 2008, 326 pp.
4. Clavel J, Mandereau L, Cordier S, et al. Hairy cell leukaemia, occupation, and smoking. *Br J Haematol.* 1995;91:154–61.
5. Monnereau A, Slager SL, Hughes AM, et al. Medical history, lifestyle, and occupational risk factors for hairy cell leukemia: the interlymph non-Hodgkin lymphoma subtypes project. *J Natl Cancer Inst Monogr.* 2014;2014:115–24.
6. Oloske D, Golomb H, Farber M, et al. A case control inquiry into the etiology of hairy cell leukemia. *Am J Epidemiol.* 1985;121:675–83.
7. Nordström M, Hardell L, Magnuson A, et al. Occupational exposures, animal exposure and smoking as risk factors for hairy cell leukaemia evaluated in a case-control study. *Br J Cancer.* 1998;77:2048–52.
8. Staines A, Cartwright RA. Hairy cell leukaemia: descriptive epidemiology and a case-control study. *Br J Haematol.* 1993;85:714–7.
9. Sopori M. Effects of cigarette smoke on the immune system. *Nat Rev Immunol.* 2002;2:372–7.
10. Oloris SCS, Frazer-Abel AA, Jubala CM, et al. Nicotine-mediated signals modulate cell death and survival of T lymphocytes. *Toxicol Appl Pharmacol.* 2010;242:299–309.
11. Bernstein L, Newton P, Ross RK. Epidemiology of hairy cell leukemia in Los Angeles County. *Cancer Res.* 1990;50:3605–9.
12. Ruiz-Arguelles GJ, Cantu-Rodriguez OG, Gomez-Almaguer D, et al. Hairy cell leukemia is infrequent in Mexico and has a geographic distribution. *Am J Hematol.* 1996;318:316–8.
13. Pascual V, Liu YJ, Magalski A, et al. Analysis of somatic mutation in five B cell subsets of human tonsil. *J Exp Med.* 1994;180:329–39.
14. Forconi F, Sahota SS, Raspadori D, et al. Hairy cell leukemia: at the crossroad of somatic mutation and isotype switch. *Blood.* 2004;104:3312–7.
15. Arons E, Sunshine J, Suntum T, et al. Somatic hypermutation and VH gene usage in hairy cell leukaemia. *Br J Haematol.* 2006;133:504–12.
16. Liu YJ, Malisan F, de Bouteiller O, et al. Within germinal centers, isotype switching of immunoglobulin genes occurs after the onset of somatic mutation. *Immunity.* 1996;4:241–50.
17. Forconi F, Sahota SS, Raspadori D, et al. Tumor cells of hairy cell leukemia express multiple clonally related immunoglobulin isotypes via RNA splicing. *Blood.* 2001;98:1174–81.
18. Arons E, Roth L, Sapolsky J, et al. Evidence of canonical somatic hypermutation in hairy cell leukemia. *Blood.* 2011;117:4844–51.
19. Basso K, Liso A, Tiacci E, et al. Gene expression profiling of hairy cell leukemia reveals a phenotype related to memory B cells with altered expression of chemokine and adhesion receptors. *J Exp Med.* 2004;199:59–68.
20. Tiacci E, Liso A, Piris M, et al. Evolving concepts in the pathogenesis of hairy-cell leukaemia. *Nat Rev Cancer.* 2006;6:437–48.
21. Forconi F, Raspadori D, Lenoci M, et al. Absence of surface CD27 distinguishes hairy cell leukemia from other leukemic B-cell malignancies. *Haematologica.* 2005;90:266–8.
22. Fecteau JF, Côté G, Nérón S. A new memory CD27-IgG+ B cell population in peripheral blood expressing VH genes with low frequency of somatic mutation. *J Immunol.* 2006;177:3728–36.
23. Vanhentenrijk V, De Wolf-Peeters C, Wlodarska I. Comparative expressed sequence hybridization studies of hairy cell leukemia show uniform expression profile and imprint of spleen signature. *Blood.* 2004;104:250–5.
24. Park JH, Tallman MS. 103 – hairy cell leukemia. 5th ed. Abeloff's Clinical Oncology, 5/e. Elsevier Inc.; 1979, 1979–1990.e4 pp.
25. Miyoshi EK, Stewart PL, Kincade PW, et al. Aberrant expression and localization of the cytoskeleton-binding pp52 (LSP1) protein in hairy cell leukemia. *Leuk Res.* 2001;25:57–67.
26. Chung SS, Kim E, Park JH, et al. Hematopoietic stem cell origin of BRAFV600E mutations in hairy cell leukemia. *Sci Transl Med.* 2014;6:238ra71.
27. Tiacci E, Trifonov V, Schiavoni G, et al. BRAF mutations in hairy-cell leukemia. *N Engl J Med.* 2011;364:2305–15.
28. Tiacci E, Schiavoni G, Martelli MP, et al. Constant activation of the RAF-MEK-ERK pathway as a diagnostic and therapeutic target in hairy cell leukemia. *Haematologica.* 2013;98:635–9.
29. Hoffman. Hairy cell leukemia. In: Hematology: basic principles and practice. 6th ed. Elsevier Inc.; 2012. p. 1192–203 [Chapter 77].
30. Grever MR. How I treat hairy cell leukemia. *Blood.* 2010;115:21–8.
31. Jain P, Pemmaraju N, Ravandi F. Update on the biology and treatment options for hairy cell leukemia. *Curr Treat Options Oncol.* 2014;15:187–209.
32. Zuzel M, Cawley J. The biology of hairy cells. *Best Pract Res Clin Haematol.* 2003;16:1–13.
33. Robak T, Matutes E, Catovsky D, et al. Hairy cell leukaemia: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2015;26:v100–7.
34. Uppal G, Ly V, Wang Z-X, et al. The utility of BRAF V600e mutation-specific antibody VE1 for the diagnosis of hairy cell leukemia. *Am J Clin Pathol.* 2015;143:120–5.
35. Cornet E, Delmer A, Feugier P, et al. Recommendations of the SFH (French Society of Haematology) for the diagnosis, treatment and follow-up of hairy cell leukaemia. *Ann Hematol.* 2014;93:1977–83.
36. Jehn U, Bartl R, Dietzfelbinger H, et al. An update: 12-year follow-up of patients with hairy cell leukemia following treatment with 2-chlorodeoxyadenosine. *Leukemia.* 2004;18:1476–81.
37. Maloisel F, Benboubker L, Gardembas M, et al. Long-term outcome with pentostatin treatment in hairy cell leukemia patients. A French retrospective study of 238 patients. *Leukemia.* 2003;17:45–51.
38. Grever MR, Zinzani PL. Long-term follow-up studies in hairy cell leukemia. *Leuk Lymphoma.* 2009;50:23–6.
39. Else M, Ruchlemer R, Osuji N, et al. Long remissions in hairy cell leukemia with purine analogs. *Cancer.* 2005;104:2442–8.
40. Giblett ER, Anderson JE, Cohen F, et al. Adenosine-deaminase deficiency in two patients with severely impaired cellular immunity. *Lancet.* 1972;300:1067–9.
41. Cohen A, Hirschhorn R, Horowitz SD, et al. Deoxyadenosine triphosphate as a potentially toxic metabolite in adenosine deaminase deficiency. *Proc Natl Acad Sci U S A.* 1978;75:472–6.
42. Johnston JB. Mechanism of action of pentostatin and cladribine in hairy cell leukemia. *Leuk Lymphoma.* 2011;52:43–5.
43. Rafel M, Cervantes F, Beltrán JM, et al. Deoxycoformycin in the treatment of patients with hairy cell leukemia: results of a Spanish collaborative study of 80 patients. *Cancer.* 2000;88:352–7.
44. Ribeiro P, Bouaffia F, Paud P, et al. Long term outcome of patients with hairy cell leukemia treated with pentostatin. *Cancer.* 1999;85:65–71.
45. Flinn IW, Kopecky KJ, Foucar MK, et al. Long-term follow-up of remission duration, mortality, and second malignancies in hairy cell leukemia patients treated with pentostatin. *Blood.* 2000;96:2981–6.
46. Grever MR, Doan CA, Kraut EH. Pentostatin in the treatment of hairy-cell leukemia. *Best Pract Res Clin Haematol.* 2003;16:91–9.
47. Steis RG, Urba WJ, Kopp WC, et al. Kinetics of recovery of CD4+ T cells in peripheral blood of deoxycoformycin-treated patients. *J Natl Cancer Inst.* 1991;83:1678–9.

48. Urba WJ, Baseler MW, Kopp WC, et al. Deoxycoformycin-induced immunosuppression with hairy cell leukemia. *Blood*. 1989;73:38–46.
49. Robak T, Blasinska-Morawiec M, Blonski J, et al. 2-Chlorodeoxyadenosine (cladribine) in the treatment of hairy cell leukemia and hairy cell leukemia variant: 7-year experience in Poland. *Eur J Haematol*. 1999;62:49–56.
50. Rosenberg JD, Burian C, Waalen J, et al. Clinical characteristics and long-term outcome of young hairy cell leukemia patients treated with cladribine: a single-institution series. *Blood*. 2014;123:177–83.
51. Goodman GR, Burian C, Koziol JA, et al. Extended follow-up of patients with hairy cell leukemia after treatment with cladribine. *J Clin Oncol*. 2003;21:891–6.
52. Chadha P, Rademaker AW, Mendiratta P, et al. Treatment of hairy cell leukemia with 2-chlorodeoxyadenosine (2-CdA): long-term follow-up of the Northwestern University experience. *Blood*. 2005;106:241–6.
53. Robak T, Błasińska-Morawiec M, Krykowski E, et al. 2-Chlorodeoxyadenosine (2-CdA) in 2-hour versus 24-hour intravenous infusion in the treatment of patients with hairy cell leukemia. *Leuk Lymphoma*. 1996;22:107–11.
54. Robak T, Jamroziak K, Gora-Tybor J, et al. Cladribine in a weekly versus daily schedule for untreated active hairy cell leukemia: final report from the Polish Adult Leukemia Group (PALG) of a prospective, randomized, multicenter trial. *Blood*. 2007;109:3672–5.
55. Lauria F, Bocchia M, Marotta G, et al. Weekly administration of 2-chlorodeoxyadenosine in patients with hairy-cell leukemia is effective and reduces infectious complications. *Haematologica*. 1999;84:22–5.
56. von Rohr A, Schmitz S-FH, Tichelli A, et al. Treatment of hairy cell leukemia with cladribine (2-chlorodeoxyadenosine) by subcutaneous bolus injection: a phase II study. *Ann Oncol*. 2002;13:1641–9.
57. Zenhäusern R, Schmitz S-FH, Solenthaler M, et al. Randomized trial of daily versus weekly administration of 2-chlorodeoxyadenosine in patients with hairy cell leukemia: a multicenter phase III trial (SAKK 32/98). *Leuk Lymphoma*. 2009;50:1501–11.
58. Nieva J, Bethel K, Saven A. Phase 2 study of rituximab in the treatment of cladribine-failed patients with hairy cell leukemia. *Blood*. 2003;102:810–3.
59. Thomas DA, Brien SO, Bueso-ramos C, et al. Rituximab in relapsed or refractory hairy cell leukemia. *Leukemia*. 2003;17:3906–11.
60. Else M, Dearden CE, Matutes E, et al. Rituximab with pentostatin or cladribine: an effective combination treatment for hairy cell leukemia after disease recurrence. *Leuk Lymphoma*. 2011;52:75–8.
61. Ravandi F, O'Brien S, Jorgensen J, et al. Phase 2 study of cladribine followed by rituximab in patients with hairy cell leukemia. *Blood*. 2011;118:3818–23.
62. Ravandi F, Jorgensen JL, O'Brien SM, et al. Eradication of minimal residual disease in hairy cell leukemia. *Blood*. 2006;107:4658–62.
63. Tacci E, Park JH, De Carolis L, et al. Targeting mutant BRAF in relapsed or refractory hairy-cell leukemia. *N Engl J Med*. 2015;373, 150909140059008.
64. Sivina M, Kreitman RJ, Arons E, et al. The bruton tyrosine kinase inhibitor ibrutinib (PCI-32765) blocks hairy cell leukaemia survival, proliferation and B cell receptor signalling: a new therapeutic approach. *Br J Haematol*. 2014;166:177–88.
65. Jain P, Polliack A, Ravandi F. Novel therapeutic options for relapsed hairy cell leukemia. *Leuk Lymphoma*. 2015;1–9.
66. Kreitman RJ, Stetler-Stevenson M, Margulies I, et al. Phase II trial of recombinant immunotoxin RFB4(dsFv)-PE38 (BL22) in patients with hairy cell leukemia. *J Clin Oncol*. 2009;27:2983–90.
67. Kreitman RJ, Tallman MS, Robak T, et al. Phase I trial of anti-CD22 recombinant immunotoxin moxatumomab pasudotox (CAT-8015 or HA22) in patients with hairy cell leukemia. *J Clin Oncol*. 2012;30:1822–8.
68. Ruiz-Delgado GJ, Tarín-Arzaga LC, Alarcón-Urdaneta C, et al. Treatment of hairy cell leukemia: long-term results in a developing country. *Hematology*. 2012;17:140–3.
69. Gómez Almaguer D, Jaime Pérez J, Herrera Garza J, et al. Interferón alfa intermitente en el tratamiento de la LCP. *Rev Invest Clin*. 1998;50:331–4.
70. Jansen JAN, Hermans JO. Splenectomy in hairy cell leukemia: a retrospective multicenter analysis. *Cancer*. 1981;47:2066–76.
71. Van Norman AS, Nagorney DM, Martin JK, et al. Splenectomy for hairy cell leukemia. A clinical review of 63 patients. *Cancer*. 1986;57:644–8.
72. Lad D, Dhawan R, Khadwal A, et al. Outcomes of splenectomy versus cladribine in resource limited settings in hairy cell leukemia. *Leuk Lymphoma*. 2014;55:1428–30.
73. Sarvaria A, Topp Z, Saven A. Current therapy and new directions in the treatment of hairy cell leukemia a review. *JAMA Oncol*. 2015;2:123–9.
74. Lopez-Rubio M, Garcia-Marco JA. Current and emerging treatment options for hairy cell leukemia. *Onco Targets Ther*. 2015;8:2147–56.
75. Poret N, Fu Q, Guihard S, et al. CD38 in hairy cell leukemia is a marker of poor prognosis and a new target for therapy. *Cancer Res*. 2015;75:3902–11.