



ORIGINAL ARTICLE

Prevalence of *Trypanosoma cruzi* infection in dogs and small mammals in Nuevo León, Mexico



Lucio Galaviz-Silva^a, Roberto Mercado-Hernández^b,
José J. Zárate-Ramos^c, Zinnia J. Molina-Garza^{a,*}

^a Universidad Autónoma de Nuevo León (UANL), Facultad de Ciencias Biológicas; Laboratorio de Patología Molecular, San Nicolás de los Garza, Nuevo León, 66451, Mexico

^b Universidad Autónoma de Nuevo León (UANL), Facultad de Ciencias Biológicas; Departamento de Ciencias Exactas y Desarrollo Humano, San Nicolás de los Garza, Nuevo León 66451, Mexico

^c Universidad Autónoma de Nuevo León (UANL), Facultad de Medicina Veterinaria y Zootecnia, Campus Ciencias Agropecuarias, Escobedo, Nuevo León, Mexico

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Abstract Chagas disease, caused by the protozoan *Trypanosoma cruzi*, is an important public health concern in areas extending from South America northward into the southern United States of America. Although this hemoflagellate has many wild and domestic mammals reported as reservoir hosts, studies on this subject are scarce in Nuevo León state, a region located in northeastern Mexico. This cross-sectional study showed that the general prevalence of *T. cruzi* infection in Nuevo León state was 14.5% (35/241), this percentage matching the ones determined by PCR and traditional diagnostics. Localities and infected mammals did not significantly differ ($\chi^2 = 6.098$, $p = 0.192$); however the number of infected animals was highly correlated with mammalian species ($p = 0.009$). Striped skunks (*Mephitis mephitis*) were found to be the most infected overall (11/34, 32.3%), while dogs (*Canis familiaris*) had the lowest prevalence. In conclusion, although the prevalence of *T. cruzi* infection in small mammals was lower in Nuevo León than in other states of Mexico, our results provide new locality records, including striped skunks, opossums (*Didelphis marsupialis*) and dogs, and extend the recorded area to woodrats (*Neotoma micropus*).

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* Corresponding author.

E-mail address: molinazinnia@hotmail.com (Z.J. Molina-Garza).

PALABRAS CLAVE

Trypanosoma cruzi;
Mamíferos
reservorios;
Zorrillos;
Perros;
Ratas de campo;
Tlacuache

Prevalencia de la infección por *Trypanosoma cruzi* en perros y pequeños mamíferos de Nuevo León, México

Resumen La enfermedad de Chagas, causada por el protozooario *Trypanosoma cruzi*, es un problema importante para la salud pública en una vasta región que se extiende en dirección norte desde Sudamérica hasta el sur de los Estados Unidos de América. Aunque este hemoflagelado tiene muchos mamíferos silvestres y domésticos reportados como reservorios, los estudios sobre este tema son escasos en el estado de Nuevo León, localizado en el noreste de México. Se efectuó un relevamiento de la prevalencia de *T. cruzi* en pequeños mamíferos cubriendo 9 municipios del estado de Nuevo León y 3 tipos de asentamiento (rural, suburbano y urbano). Se observó una prevalencia general de la infección del 14,5% (35/241) usando PCR y diagnóstico tradicional para detectarla. No se determinó una asociación estadísticamente significativa entre las localidades relevadas y las especies de mamíferos infectados ($\chi^2 = 6,098$, $p = 0,192$); sin embargo, el número de animales infectados se correlacionó con la especie de mamífero ($p = 0,009$). La mayor prevalencia de *T. cruzi* se detectó en los zorrillos (*Mephitis mephitis*) (11/34; 32,3%); la menor (13/136; 9,5%), en los perros (*Canis familiaris*). La prevalencia de la infección por *T. cruzi* en pequeños mamíferos fue más baja en Nuevo León que en otros estados de México. Estos resultados proveen nuevos registros de localidad incluyendo zorrillos, zarigüeyas o tlacuaches (*Didelphis marsupialis*) y perros, y también amplían el área registrada para las ratas de campo (*Neotoma micropus*).

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Introduction

Chagas disease, caused by the hemoflagellate parasite *Trypanosoma cruzi*, is a widespread and endemic parasitosis extending from South America northward into the southern United States of America. It is currently a major cause of morbidity and mortality, with 6–7 million people infected and approximately 21 000 deaths per year³². *Trypanosoma cruzi* is a multihost parasite that affects more than 180 mammalian species; it displays huge intraspecific heterogeneity and a complex transmission cycle, which may exhibit local peculiarities that occur because the distribution of *T. cruzi* infection is not homogeneous among houses, localities, and/or biomes²⁵.

In northeastern (NE) Mexico, including Nuevo León state, reports of wild and domestic reservoir hosts of *T. cruzi* are deficient. In contrast, with numerous descriptions from southern and central Mexico of wild mammals of epidemiological importance including nine-banded armadillos (*Dasypus novemcinctus*), opossums (*Didelphis marsupialis* and *D. virginiana*), rodents (*Sigmodon hispidus*, *Mus musculus*), white-nosed coati (*Nasua narica*) and raccoon (*Procyon lotor*)^{16,30,31}. Moreover, dogs (*Canis familiaris*) are considered the predominant domestic reservoir hosts for *T. cruzi* in many areas of endemicity. This infection has been poorly studied in Mexico, with some dispersed studies in Jalisco, Mexico, Morelos, Puebla, and Yucatán^{6,9,14,23,28}. Furthermore, some records have been published from southeastern USA (Louisiana, Oklahoma, Georgia and Texas) where the epidemiological role of domestic dogs as urban and rural mammalian hosts of *T. cruzi* is becoming increasingly important, as indicated by their high seroprevalence^{3,13}; despite this condition (of high seroprevalence) all dogs display low

parasitemia, and no hemoculture has been positive for the parasite in some municipalities of Brazil³³.

Northeastern Mexico shares two items with southeastern (SE) USA (Texas and New Mexico): a border and an epidemiological profile, in which peridomestic and wild triatomines, *Triatoma gerstaeckeri* (Stål), are associated with wild mammals^{7,18}. However, in NE Mexico only the Southern plains woodrat (*Neotoma micropus*) has been reported¹¹. The objective of this study was to assess the prevalence of *T. cruzi* in domestic canines and wild small mammals in Nuevo León, based on the responses to previous questionnaires regarding vector recognition and domestic dogs from inhabitants of this state¹⁹, where they have been reported as an important risk factor when they are infected with *T. cruzi* and live in close contact with the human population.

Materials and methods

Study area

Nuevo León state is located in the foothills of the Sierra Madre Oriental mountain range (27°49'N, 23°11'S and 98°26'E, 101°14'W). This study was conducted from March 2011 to November 2013 in nine municipalities, which correspond to: a) a rural area comprising General Terán, Dr. Arroyo and Cerralvo; b) a suburban county located in Allende; and c) an urban area comprising Guadalupe, San Nicolás de los Garza, Escobedo, Apodaca and Monterrey, the Capital of the state (Fig. 1). These localities were selected according to a recent report by Molina-Garza et al¹⁹.

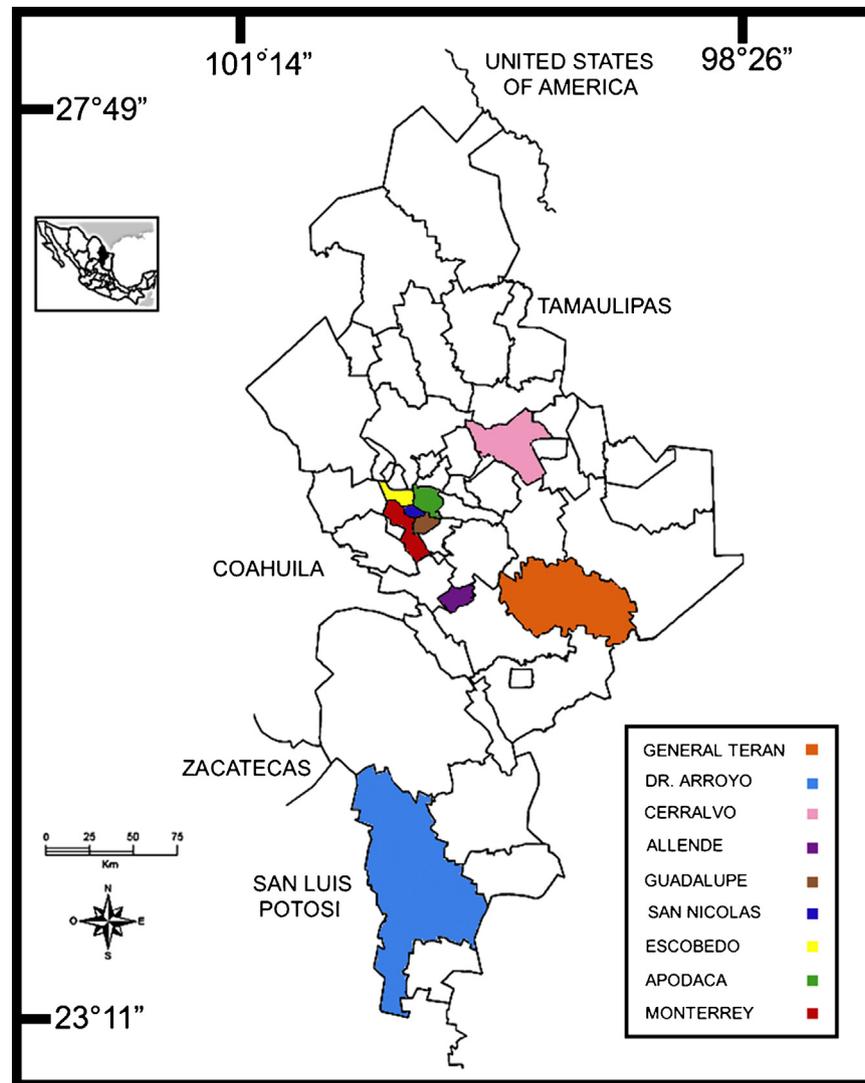


Figure 1 Map of Nuevo León showing the sampled localities; a rural zone (General Terán, Dr. Arroyo and Cerralvo); a suburban county (Allende) and an urban area (Apodaca, Escobedo, Guadalupe, Monterrey, and San Nicolás).

Study design. Capture and handling of small wild mammals

This is a cross sectional study. Wild mammal hosts were collected using live traps (H.B. Sherman Traps, Tallahassee, FL, and Havahart Traps, Litzitz, PA) baited with peanut butter and/or sardines during 5 days per month in each municipality to perform a probabilistic sample stratified in three different localities, rural, suburban and urban, where the number of samples equally represented in a sample size calculated among 204–322 individuals, based in an estimated prevalence of *T. cruzi* infection of 15.8–30% reported in Mexican states^{6,9}, with an absolute precision of 1.96, a confidence level of 95%, and $E = 5\%$ ¹⁹.

Fifty traps were placed at an interval of 2-m each in four linear transects of 100 m each during the afternoon. Another fifty traps were specifically set for *N. micropus*, following the track of cacti (locally known as “nopaleras”, *Opuntia* spp.), pads bitten by the rodents and typical stool pellets found at the base of nests built out of dry sticks⁷. The total

capture effort was standardized to 250traps/night. Traps were set during the afternoon and checked the following morning, as specified by the guidelines of the Mexican Secretary of Environment and Natural Resources²⁰. Captured animals were anesthetized by an intramuscular injection of ketamine (100 mg/kg of body weight; Fort Dodge Laboratories, Inc., Fort Dodge, IA, USA)¹². Woodrats were euthanized in cervical dislocation and skunks and opossums by intracardiac injection with an overdose of sodium pentobarbital (Butler Company, Columbus, OH), followed by exsanguination⁷. Small mammals were taxonomically identified, classified as juveniles or adults based on weight and tooth wear, and sexed during sample collection by personnel of the Department of Zoology, UANL.

Dog surveys

Dogs were sampled in the presence and with the consent of their owners during domiciliary visits and at veterinary clinics and hospitals. Between two and 10 housing blocks were

Table 1 The prevalence of *Trypanosoma cruzi* infection among different study species of wild mammals and dogs in Nuevo León, México

Mammals	Rural +/n ^a (%)	Suburban	Urban	+/n (%) ^b	F ^c (95% CI) ^d	LM ^e (%)	PCR (%)
<i>Didelphis marsupialis</i>	2/13 (15.4)	1/8 (12.5)	3/15 (20)	6/36 (16.7)	0.167 (0.066–0.326)	3 (50)	6 (100)
<i>Neotoma micropus</i>	4/25 (16)	1/10 (10)	–	5/35 (14.3)	0.143 (0.027–0.259)	2 (40)	5 (100)
<i>Mephitis mephitis</i>	5/15 (33.3)	2/5 (40)	4/14 (28.5)	11/34 (32.3)	0.323 (0.168–0.484)	5 (45.4)	10 (90.9)
<i>Canis familiaris</i>	9/58 (15.5)	3/27 (11.1)	1/51 (1.9)	13/136 (9.5)	0.095 (0.046–0.144)	5 (38.5)	10 (76.9)
Total	20/111 (18)	7/50 (14)	8/80 (10)	35/241 (14.5)	0.145 (0.101–0.189)	15 (42.9)	31 (88.6)

Number of infected animals and species with a significance of $\chi^2 = 11.5$, $df = 3$, $p = 0.009$.

^a Sample size by habitat.

^b Positives/n total by species (prevalence).

^c Frequency of positive animals.

^d 95% confidence interval (CI).

^e Light microscopy (blood smears or histopathology).

selected for sampling from maps, according to the density of each locality (rural, suburban or urban). The selection procedure for the houses to be sampled involved the enumeration of each house in the block, and the selection of five houses on each side, with the previous consent of the householder. Participation in the study was random and voluntary, with the sample size required described above.

A questionnaire completed by the dogs' owners and veterinarians ascertained information regarding birthplace, sex, age (puppies = 1–4 months; young or juvenile dogs = 6–24 months; adult dogs = >2 years) and general body condition². Several of the dogs were found admitted to veterinary hospitals, and in the case of death, organ donation was requested.

Sample collection and parasitological diagnosis

Blood samples from both small mammals and dogs were collected in EDTA vacutainer tubes (Beckton, Dickinson and Company, Franklin Lakes, NJ, USA) by puncturing the cephalic vein. Blood smears were prepared directly *in situ* with freshly sampled blood; smears were stained with Giemsa stain. The remaining blood samples were stored at -20°C for PCR²². Tissue samples (heart, skeletal muscle, smooth muscle, liver, spleen, lung, kidney, pancreas and brain) from each necropsied wild animal were fixed in 10% buffered formalin for processing. Routine histopathology was carried out using the hematoxylin and eosin (H & E) staining technique and the samples were closely examined for the presence of amastigote nests⁷. Representative photomicrographs were digitized using a Leica photomicroscope linked to a DFC 480 digital camera (Image Driving Software, LW Scientific, Inc., Tucker, GA, USA).

Identification of *T. cruzi* by PCR

Total DNA was extracted from whole blood using the standard DNAzol technique following the manufacturer's instructions (Invitrogen, San Diego, CA, USA). The extracted DNA pellet was resuspended in 50 μl of deionized sterile water and stored at -20°C until use. Presence of *T. cruzi* DNA was confirmed by amplifying a region of the kinetoplastid minicircle (kDNA), using the oligonucleotide primers

KNS1 (5'-GGG GTT CG A TTG GGG TTG GTG TA-3') and KNS2 (5'-AAA (G/T)TT GAA CGC CCC TCC CAA A-3'), annealing in the conserved microregion of the minicircles and yielded products of 310 bp, as described elsewhere^{1,18}.

Statistical analysis

Reservoirs were considered infected by *T. cruzi* when one traditional parasitological diagnostic or PCR was positive⁸. Comparisons of the trypanosome infection rates for different age classes, sex and species of small mammals and dogs by communities (rural, suburban, and urban) were calculated using 2×2 contingency tables and the chi-squared (χ^2) or the Fisher's exact test when appropriate. Data were analyzed using SPSS software, version 17 (Chicago, IL).

Results

A total of 241 mammals from four different species were analyzed, included 136 dogs (117 mixed-breed dogs and 19 purebreds, 86.0% and 13.9% respectively), 36 opossums (*D. marsupialis*), 35 woodrats (*N. micropus*) and 34 striped skunks (*M. mephitis*). The number of infected animals highly differed between the species ($\chi^2 = 11.54$, degree freedom [df] = 3, $p = 0.009$). Striped skunks were found to be the most infected wild reservoir hosts in all the different localities (11/34, 32.3%, confidence intervals [CI] = 0.168–0.484), followed by opossums (16.7%, CI = 0.066–0.326) and woodrats (14.3%, CI = 0.027–0.259, Table 1).

With regard to habitats, one hundred eleven mammalian hosts were analyzed from rural municipalities, 50 from suburban areas, and 80 from urban municipalities (Table 1). The capture success prevalence for the different localities was 8.8%, 4% and 6.4%, respectively, and was highly associated by area and the number of small mammals species ($\chi^2 = 30.5$, $df = 2$, $p = 0.002$). A total of 35 animals (35/241, 14.5%) were found infected with *T. cruzi* (Fig. 2A), 20 from rural (the highest prevalence, 20/111, 18%), seven from suburban (14%) and eight from urban (10%) localities. Although the infection prevalence by species of mammal reservoir hosts did not significantly differ between the localities ($\chi^2 = 6.09$, $df = 4$, $p = 0.192$), the infection prevalence by locality and sex was highly dependent ($\chi^2 = 206.5$, $df = 12$, $p = 0.000$). The

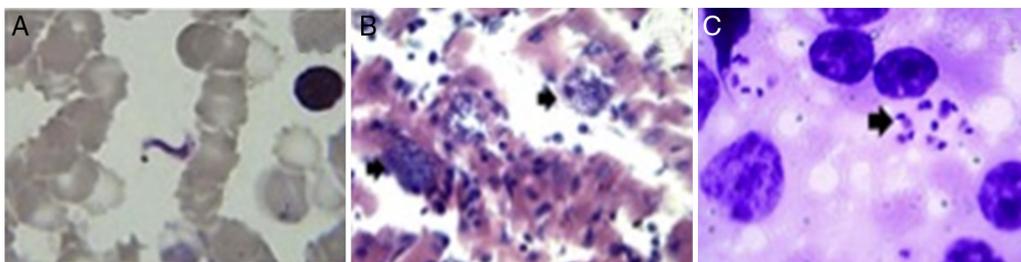


Figure 2 *Trypanosoma cruzi* from wild mammals and dogs. (A) Giemsa stained blood smear from an opossum showing trypomastigotes of *Trypanosoma cruzi*. (B) Histopathological lesions with amastigote nests in the cardiac muscle of *T. cruzi* infected dogs (arrow indicates amastigote nests). (C) Amastigote forms (arrow) of *T. cruzi* observed in two mixed-breed puppies.

highest prevalence of infected animals were observed in striped skunks from rural (33.3%), suburban (40%) and urban areas (28.5%), whereas dogs showed the lowest prevalence in these three habitats (Table 1).

The prevalence of *T. cruzi* infection by age, in suburban area was the highest in juvenile striped skunks (100%) followed by juvenile opossums (50%). There is statistical dependence among infection rate, age class and animal species ($\chi^2 = 47.8$; $df = 15$, $p = 0.000$). With regard to dog infection by age, in rural areas juveniles reached the highest (35.7%, 5/14) followed by adults (13.3%, 2/15) and juveniles (12.5%, 1/8) in the suburban area; only one male adult dog was found to be infected (3.7%, 1/27) in the urban area (Table 2).

With regard to the sex of the sampled animals, 134 were males (55.6%) and 107 females (44.4%). Twenty females tested positive (18.7%), indicating a higher prevalence of infection with *T. cruzi* in females than in males

(15/134, 11.2%). However this difference was not significant ($\chi^2 = 0.155$, $df = 3$, $p = 0.984$). Furthermore, the prevalence of infection was highest in female striped skunks (50% in the suburban area), followed by male in the rural region (40%); however, the analyses among prevalence of infection, species of mammalian reservoir hosts and sex were significant ($\chi^2 = 15.32$, $df = 6$, $p = 0.018$) (Table 2).

Questionnaire results

Clinical signs suggestive of infection reported by pet owners were confirmed microscopically and by molecular diagnostics. One mixed-breed puppy from the rural area (Cerralvo) and one adult from the metropolitan area showed lethargy, anorexia and fever. Gross necropsy indicated that *T. cruzi* infection was preferentially located in the adipose tissue, skeletal muscle and cardiac muscle (Fig. 2B). There was

Table 2 The prevalence of infection by *T. cruzi* in the small mammalian fauna and dogs from Nuevo León, México by sex and age classes

Mammals	Sample	Sex		Age classes (n/+)		
		Male	Female	Adult	Juvenile	Puppies
<i>Rural area</i>						
<i>Didelphis marsupialis</i>	13	14.3 (1/7)	16.7 (1/6)	16.7 (1/6)	14.3 (1/7)	-
<i>Neotoma micropus</i>	25	6.2 (1/16)	33.3 (3/9)	19.0 (4/21)	0 (0/4)	-
<i>Mephitis mephitis</i>	15	40 (4/10)	20 (1/5)	33.3 (4/12)	33.3 (1/3)	-
<i>Canis familiaris</i>	58	14.8 (4/27)	16.1 (5/31)	8.3 (2/24)	35.7 (5/14)	10 (2/20)
Subtotal	111	16.6 (10/60)	19.6 (10/51)	17.4 (11/63)	25 (7/28)	10 (2/20)
<i>Suburban area</i>						
<i>Didelphis marsupialis</i>	8	0 (0/5)	33.3 (1/3)	(0/6)	50 (1/2)	-
<i>Neotoma micropus</i>	10	25 (1/4)	0 (0/6)	14.3 (1/7)	0 (0/3)	-
<i>Canis familiaris</i>	3/27	0 (0/15)	25 (3/12)	13.3 (2/15)	12.5 (1/8)	0 (0/4)
<i>Mephitis mephitis</i>	2/5	33.3 (1/3)	50 (1/2)	25 (1/4)	100 (1/1)	-
Subtotal	50	7.4 (2/27)	21.7 (5/23)	12.5 (4/32)	21.4 (3/14)	0 (0/4)
<i>Urban area</i>						
<i>Didelphis marsupialis</i>	15	14.3 (1/7)	25 (2/8)	27.2 (3/11)	0 (0/4)	-
<i>Canis familiaris</i>	51	2.9 (1/34)	0 (0/17)	3.7 (1/27)	0 (0/16)	0 (0/8)
<i>Mephitis mephitis</i>	14	1.6 (1/6)	37.5 (3/8)	33.3 (4/12)	0 (0/2)	-
Subtotal	80	6.3 (3/47)	15.1 (5/33)	16 (8/50)	0 (0/22)	0 (0/8)
Total	241	11.2 (15/134)	18.7 (20/107)	15.9 (23/145)	25.6 (10/64)	6.2 (2/32)

n = sample size by mammalian species. + = positive.

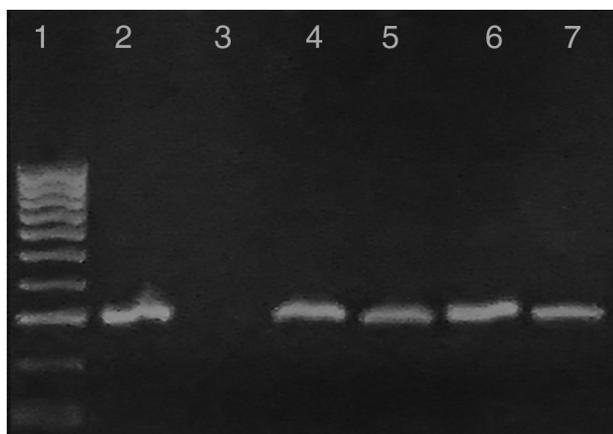


Figure 3 Characterization of kDNA-PCR-positive samples from infected reservoirs: Lane 1: 100-bp DNA ladder; 2: *T. cruzi* positive control; 3: negative control; 4 and 5: representative amplicons from DNA extracted from a *T. cruzi*-infected adult dog and a mixed-breed puppy in acute phase from metropolitan and rural areas, respectively; 6: representative sample of positive PCR from woodrats; 7: PCR product from a striped skunk.

a high prevalence of intracellular amastigote nests and interstitial inflammation, accompanied by perivascularitis and adipose degeneration. The kidney and spleen (Fig. 2C) had moderate sinusoidal congestion; however, no pathological changes were found in the brain, lung and pancreas and the parasite did not appear to be present in these tissues. The diagnosis was confirmed by PCR (Fig. 3) and a few weeks later they both succumbed to the infection, showing evidence of the classical histopathological lesions (Fig. 2B).

Discussion

This study provided clear evidence of the active cycles of *T. cruzi* transmission in NE Mexico and demonstrated the first autochthonous cases in the state of Nuevo León, with direct microscopic examination and PCR in striped skunks, dogs and opossums. Therefore, the dogs can be used as sentinels of the local transmission of Chagas disease; the regular screening of domestic dog populations can be used to identify houses or clusters with high risk of transmission^{5,25}. We have reported two acute cases (dogs from Cerralvo and urban area) of infection among 13 positive dogs with moderate to high parasitemia levels; these findings support the reports from USA (Texas) and Argentina (Chaco), where the most frequent clinical signs were an enlarged heart, lethargy and anorexia in acute infection and death in puppies younger than one year old^{5,13,21}, suggesting a fatal course of infection in the dogs sampled, similarly to those reported in cases from Colombia²⁹.

The distribution of *T. cruzi* infection was not homogeneous among municipalities, *i.e.*, the highest infection prevalence was observed in dogs from rural areas, followed by the suburban and urban areas of Nuevo León, where the prevalence varied from 1.9% to 15.5%. This profile was also observed in Brazilian biomes (the Amazon, Caatinga, and Pantanal), where the distribution of *T. cruzi* infection ranged from 0%–42% to 11%–89%^{24,33}. Previous reports also identified domestic pets as an important risk factor

associated with Chagas disease in human populations from Toluca, Puebla and Morelos, Mexico^{9,23,28}; however, the most serious infection prevalence in dogs was documented in Texas, USA, with 20% PCR-positive cases¹³, suggesting that the infection is steadily maintained in these reservoir hosts and opportunistic vertebrates (opossums, striped skunks and woodrats), which are used to live in close proximity to human dwellings³. Our results involved direct microscopic examinations (blood smears and histopathology) and PCR assays, showing a percentage of matches between the two analyses of 42.9 and 88.6% respectively (Table 1). Moreover, results of 11–22% with parasitological methods (hemoculture, xenodiagnosis and xenoculture), 67–100% by PCR have been reported¹, showing lower levels of prevalence than those reported with serological tests, which yielded infection rates of up to 100%^{24,25}. However, a high association between parasitological and serological results has been reported. Furthermore, PCR^{1,14} allows to avoid some cases of false positive or false negative results, which could rise from *Leishmania* spp. or *T. rangeli*⁸.

A difference between our study area and the Amazon region is observed. In Brazil dogs have an elevated prevalence of contact with the wild environment, because houses are practically located inside wild forest areas, and it is difficult to delimit peridomestic and wild areas, and dogs are involved in hunting activities²⁴, whereas in NE Mexico the scenario is completely different, because human residences are built after the destruction of the wild environment. This could be the cause that our results did not show any correlation between *T. cruzi* infection in dogs and small wild mammals as described³³.

When we examined the prevalence of infection in dogs by age, the age-prevalence curve increased from 10% in puppies to 35.7% in juvenile dogs, which had the highest *T. cruzi* infection rate. A similar scenario is described in Panama²⁷, which highlights the importance of puppies and juvenile dogs in the continuous life cycle of *T. cruzi* in the rural region of Nuevo León and also the epidemiological importance of the single positive adult recorded in an urban area. This dog was involved in camping activities around the mountains close to the state of Nuevo León, recognized as a non-endemic region for *T. cruzi* infection¹⁰, which indeed does not have a Chagas disease vector control program such as those existing in some South American countries.

Furthermore, there are no records of *T. cruzi* infection prevalence in striped skunks in Mexico, and no systematic studies of its occurrence in small wild mammals. In this study, the highest prevalence of infection was found in the striped skunk in suburban, rural and urban areas, suggesting that this species is an important host for *T. cruzi* infection, as has been demonstrated in the southeast of the USA⁴. Several studies have reported the role of opossums as natural wild reservoir hosts of *T. cruzi* in dwellings and peridomestic areas in Chiapas, Yucatán, Jalisco and Mexico state^{15,26,30,31}. In the southern United States and Argentina, the opossum is considered to be the main wild mammal host species as a synanthropic reservoir^{4,7,21}; nevertheless our results reported opossums as the second most important wild reservoir host for *T. cruzi*. In this study, opossums and striped skunks were collected close to highways near a metropolitan area that had grown substantially in the last 10 years. As a result of the deforestation caused by the building of

new residential areas, these vertebrate reservoirs have been strongly forced to visit peridomestic habitats in search of food and to increase the interaction between the sylvatic and domestic cycles; the consequence of this process is the increased opportunity for contact among humans, domestic animals and wildlife³³.

Regarding woodrats, previous studies in General Terán, NL (a rural county) reported that the prevalence of *T. cruzi* infection had been lower than in this study¹¹; the triatomine *T. gerstaeckeri*, has been found to be associated to *N. micropus* nets naturally infected with *T. cruzi* (33%–59.6%)^{17,18}, although some reports have described *T. gerstaeckeri* as a human and livestock pest species because the adults are frequent invaders of rural houses in southwestern USA³, highlighting their importance by their vectorial capacity in this region. Although the genotyping of *T. cruzi* was not performed in this work, other investigations have documented different *T. cruzi* strains in the reservoirs described, the most abundant being TcI, which is distributed by triatomine vectors and is associated with sylvatic and domestic cycles³⁴.

In conclusion, although the prevalence of *T. cruzi* infection was lower in reservoirs in Nuevo León than in other states of Mexico, this plays an important role in the maintenance and transmission cycles of *T. cruzi* due to the close proximity of the reservoir species with human domiciles. The results of this study provide new locality records for striped skunks, opossums and dogs in rural, suburban, and urban counties in NE Mexico and extend the localities reported to the woodrat previously recorded only in General Terán¹⁸ to other rural and suburban counties including Dr. Arroyo, Cerralvo and Allende. It would be necessary to consider all the results obtained and complemented with other epidemiological reports to classify Nuevo León state from a non-endemic to an endemic region, since our current and previous results support this change^{10,19}.

Ethical statement

All procedures involving animals were evaluated and approved by the Ethical Principles of Animal Experimentation established by the Committee of Ethical of Animal Experimentation of the Universidad Autónoma de Nuevo León, in agreement with the international ethical standards of the American Society of Mammalogists¹².

Ethical responsibilities

Protection of people and animals. The authors state that the procedures followed conformed to the ethical standards of the responsible human experimentation committee and in agreement with the World Medical Association and the Declaration of Helsinki.

Confidentiality of data. The authors state that no patient data appears in this article.

Right to privacy and informed consent. The authors state that in this article there are no patient data

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Conflicts of interest

The authors declare that they have no conflicts of interest

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References

1. Araújo FMGD, Bahia MT, Magalhães NMD, Martins-Filho OA, Veloso V, Tafuri WL, Gavao LMG, Carneiro CM, de Lana M, Chiari E, Soares KA, Bahia MT. Follow-up of experimental chronic Chagas' disease in dogs: use of polymerase chain reaction (PCR) compared with parasitological and serological methods. *Acta Trop.* 2002;81:21–31.
2. Baldwin K, Bartges J, Buffington T, Freeman LM, Grabow M, Legred J, Ostwald D. AAHA nutritional assessment guidelines for dogs and cats. *J Am Anim Hosp Assoc.* 2010;46:285–96.
3. Beard CB, Pye G, Steurer FJ, Rodríguez R, Campman R, Peterson AT, Ramsey J, Wirtz RA, Robinson LE. Chagas disease in a domestic transmission cycle in Southern Texas, USA. *Emerg Infect Dis.* 2003;9:103–5.
4. Brown EL, Roellig DM, Gompper ME, Monello RJ, Wenning KM, Gabriel MW, Yabsley MJ. Seroprevalence of *Trypanosoma cruzi* among eleven potential reservoir species from six states across the southern United States. *Vector-Borne Zoonotic Dis.* 2010;10:757–63.
5. Cardinal MV, Lauricella MA, Marcet PL, Orozco MM, Kitron U, Gürtler RE. Impact of community-based vector control on house infestation and *Trypanosoma cruzi* infection in *Triatoma infestans*, dogs and cats in the Argentine Chaco. *Acta Trop.* 2007;103:201–11.
6. Carrillo-Peraza JR, Manrique-Saide P, Rodríguez-Buenfil JC, Escobedo-Ortegón JF, Rodríguez-Vivas RI, Bolio-González ME, Barrera-Pérez M, Reyes-Novelo E, Sauri-Arceo CH. Estudio serológico de la tripanosomiasis Americana y factores asociados en perros de una comunidad rural de Yucatán, México. *Arch Med Vet.* 2014;46:75–81.
7. Charles RA, Kjos S, Ellis AE, Barnes JC, Yabsley MJ. Southern plains woodrats (*Neotoma micropus*) from Southern Texas are important reservoirs of two genotypes of *Trypanosoma cruzi* and host of a putative novel *Trypanosoma* species. *Vector-Borne Zoonotic Dis.* 2013;13:22–30.
8. Crisante G, Rojas A, Teixeira MM, Añez N. Infected dogs as a risk factor in the transmission of human *Trypanosoma cruzi* infection in western Venezuela. *Acta Trop.* 2006;98:247–54.
9. Estrada-Franco JG, Bhatia V, Diaz-Albiter H, Ochoa-Garcia L, Barbabosa A, Vazquez-Chagoyan JC, Martinez-Perez MA, Guzman-Bracho C, Garg N. Human *Trypanosoma cruzi* infection and seropositivity in dogs, Mexico. *Emerg Infect Dis.* 2006;12:624–30.

10. Galaviz-Silva L, Molina-Garza DP, González-Santos MA, Mercado-Hernández R, González-Galaviz JR, Molina-Garza ZJ. Update on seroprevalence of anti-*Trypanosoma cruzi* antibodies among blood donors in Northeast Mexico. *Am J Trop Med Hyg.* 2009;81:404–40.
11. Galaviz-Silva L, Arredondo JM. First report on *Neotoma micropus* (Rodentia) as a reservoir of *Trypanosoma cruzi* in Mexico. *Bol Chil Parasitol.* 1992;47:54–7.
12. Gannon WL, Sikes RS. Guidelines of the American Society of Mammalogists for the use of wild mammals in research. *J Mammal.* 2007;88:809–23.
13. Kjos SA, Snowden KF, Craig TM, Lewis B, Ronald N, Olson JK. Distribution and characterization of canine Chagas disease in Texas. *Vet Parasitol.* 2008;152:249–56.
14. Jiménez-Coello M, Acosta-Viana KY, Guzman-Marin E, Gomez-Rios A, Ortega-Pacheco A. Epidemiological survey of *Trypanosoma cruzi* infection in domestic owned cats from the tropical southeast of Mexico. *Zoonoses Public Health.* 2012;59:102–9.
15. Martínez I, Martínez-Ibarra A, Arce-Fonseca M, Rodríguez-Morales O, Pérez-Morales D, Reyes-López PA, Espinoza B. Seroprevalence and major antigens recognized by sera from *Trypanosoma cruzi*-infected dogs from Jalisco, México. *Rev Argent Microbiol.* 2014;46:85–90.
16. Martínez-Hernandez F, Rendon-Franco E, Gama-Campillo L, Villanueva-García C, Romero-Valdovinos M, Maravilla P, Alejandro-Aguilar R, Rivas N, Córdoba-Aguilar A, Muñoz-García CI, Villalobos G. Follow-up of natural infection with *Trypanosoma cruzi* in two mammals species, *Nasua narica* and *Procyon lotor* (Carnivora: Procyonidae): evidence of infection control. *Parasit Vectors.* 2015;7:405, <http://dx.doi.org/10.1186/1756-3305-7-405>
17. Martínez-Ibarra JA, Galaviz-Silva L, Lara-Campos C, Trujillo JC. Distribución de los triatominos asociados al domicilio humano en el municipio de General Terán, Nuevo León, México. *Southwest Entomol.* 1991;17:261–5.
18. Molina-Garza ZJ, Rosales-Encina JL, Galaviz-Silva L, Molina-Garza D. Prevalencia de *Trypanosoma cruzi* en triatominos silvestres de Nuevo León, México. *Salud Publica Mex.* 2007;49:37–44.
19. Molina-Garza ZJ, Rosales-Encina JL, Mercado-Hernández R, Molina-Garza DP, Gomez-Flores R, Galaviz-Silva L. Association of *Trypanosoma cruzi* infection with risk factors and electrocardiographic abnormalities in northeast Mexico. *BMC Infect Dis.* 2014;14:117, <http://dx.doi.org/10.1186/1471-2334-14-117>
20. Norma Oficial Mexicana. NOM-059-SEMARNAT. Protección ambiental-Especies nativas de México de flora y fauna silvestres-Categorías de riesgo y especificaciones para su inclusión, exclusión o cambio-Lista de especies en riesgo. *Diario Oficial de la Federación.* 2010;2:1–78.
21. Orozco MM, Enriquez G, Alvarado-Otegui JA, Cardinal V, Schijman AG, Kitron U, Gürtler RE. New sylvatic hosts of *Trypanosoma cruzi* and their reservoir competence in the humid Chaco of Argentina: a longitudinal study. *Am J Trop Med Hyg.* 2013;88:872–82.
22. Pineda V, Saldaña A, Monfante I, Santamaria A, Gottdenker NL, Yabsley MJ, Rapoport G, Calzada JE. Prevalence of trypanosome infections in dogs from Chagas disease endemic regions in Panama, Central America. *Vet Parasitol.* 2011;178:360–3.
23. Portugal-García C, García-Vázquez Z, Monteón-Padilla V, Chávez-López V, Olamendi-Portugal M, Ramos C. Anticuerpos contra *Trypanosoma cruzi* en humanos y perros y presencia del parásito en *Meccus pallidipennis* en la localidad de Puente Panitlán, Morelos, México. *Biomedica.* 2011;22:67–75.
24. Rocha FL, Roque AL, Arrais RC, Santos JP, Lima VS, Xavier SC, Cordeir-Estrella P, DiAndrea PS, Jansen AM. *Trypanosoma cruzi* TcI and TcII transmission among wild carnivores, small mammals and dogs in a conservation unit and surrounding areas, Brazil. *Parasitology.* 2013;140:160–70.
25. Roque ALR, Xavier SC, da Rocha MG, Duarte AM, D'Andrea PS, Jansen AM. *Trypanosoma cruzi* transmission cycle among wild and domestic mammals in three areas of orally transmitted Chagas disease outbreaks. *Am J Trop Med Hyg.* 2008;79:742–9.
26. Ruiz-Piña H, Cruz-Reyes A. The opossum *Didelphys virginiana* as a synanthropic reservoir of *Trypanosoma cruzi* in Dzidzilché, Yucatán, México. *Mem Inst Oswaldo Cruz.* 2002;95:613–20.
27. Saldaña A, Calzada JE, Pineda V, Perea M, Rigg C, González K, Santamaria AM, Gottdenker NL, Chaves LF. Risk factors associated with *Trypanosoma cruzi* exposure in domestic dogs from a rural community in Panama. *Mem Inst Oswaldo Cruz.* 2015;110:936–44.
28. Sosa-Jurado F, Zumaquero-Ríos JL, Reyes PA, Cruz-García A, Guzmán-Bracho C, Monteón VM. Factores bióticos y abióticos que determinan la seroprevalencia de anticuerpos contra *Trypanosomacruzi* en el municipio de Palmar de Bravo, Puebla, México. *Salud Publica Mex.* 2004;46:39–48.
29. Turriago GBC, Vallejo GA, Guhl F. Seroprevalencia de *Trypanosoma cruzi* en perros de dos áreas endémicas de Colombia. *Revista Med.* 2003;16:11–8.
30. Villagran ME, Martínez-Ibarra JA, Diego A. Pathological alterations and prevalence of *Trypanosoma cruzi* in opossums from western Mexico. *Bol Mal Salud Amb.* 2011;51:87–8.
31. Villegas-García JC, Santillán-Alarcon S. American trypanosomiasis in central Mexico: *Trypanosoma cruzi* infection in triatomine bugs and mammals from the municipality of Jiutepec in the state of Morelos. *Ann Trop Med Parasitol.* 2004;98:529–32.
32. WHO (World Health Organization). Chagas disease (*American trypanosomiasis*). In: Fact Sheet No 340 [updated 02.07.16]; 2014. Available from: <http://www.who.int/mediacentre/factsheets/fs340/es/> [accessed 15.09.16].
33. Xavier SC, Roque ALR, Lima VS, Monteiro KJ, Otaviano JCR, Ferreira Silva LF, Jansen AM. Lower richness of small wild mammal species and Chagas disease risk. *PLoS Negl Trop Dis.* 2012;6:e1647, <http://dx.doi.org/10.1371/journal.pntd.0001647>
34. Zingales B, Miles MA, Campbell DA, Tibayrenc M, Macedo AM, Teixeira MM, Schijman AG, Llewellyn MS, Lages-Silva E, Machado CR, Andrade SG, Sturm NR. The revised *Trypanosoma cruzi* subspecific nomenclature: rationale, epidemiological relevance and research applications. *Infect Genet Evol.* 2012;12:240–53.